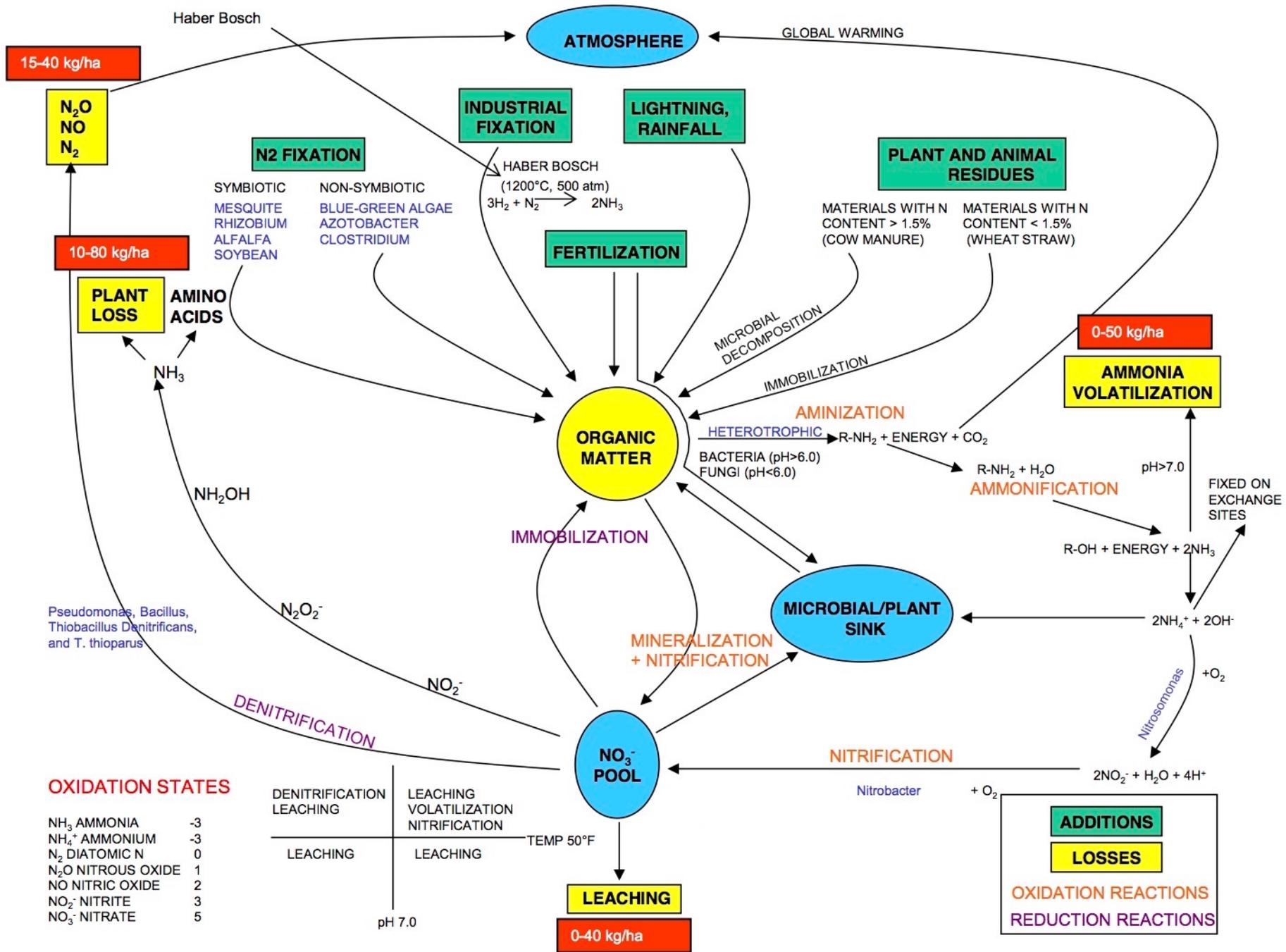


Adventist Agricultural Association  
4<sup>th</sup> Annual Convention  
2018  
Session 1

# Nitrogen Cycle

PRESENTED BY:

MICHAEL ROCKY TREVIZO



# Forms of soil N

- Elemental ( $N_2$ )
  - Gas in soil atmosphere
  - Dissolved in soil water
  - Symbiotic and non-symbiotic fixation
    - Generates  $NH_4^+$
- Inorganic forms
  - Gases –present in small amounts in soils
    - $N_2O$ ,  $NO$ ,  $NO_2$ ,  $NH_3$
  - Ionic forms
    - $NH_4^+$  -exchangeable and non-exchangeable forms
    - $NO_3^-$ ,  $NO_2^-$
  - Organic forms -98% of total soil N

# N Transformations and cycling in soils

N Gains	N losses	N Cycling (within soil, N not lost)
Fixation (biological or mechanically – fertilizer)	Plant Uptake	Immobilization
Animal manure	Denitrification	Mineralization
Crop residue	Volitization	Nitrification
	Leaching	
	Ammonium fixation	

# Vocabulary

- Nitrification: Conversion of  $\text{NO}_2$  to  $\text{NO}_3$
- Denitrification: Conversion of  $\text{NO}_3$  to N gas
- Immobilization: Conversion of PAN to Plant unavailable N
- Nitrogen Fixation: Non Organic/Non Mineral forms of N to Organic forms of N
- Mineralization: The conversion of an element from an organic form to an inorganic state as a result of microbial decomposition.

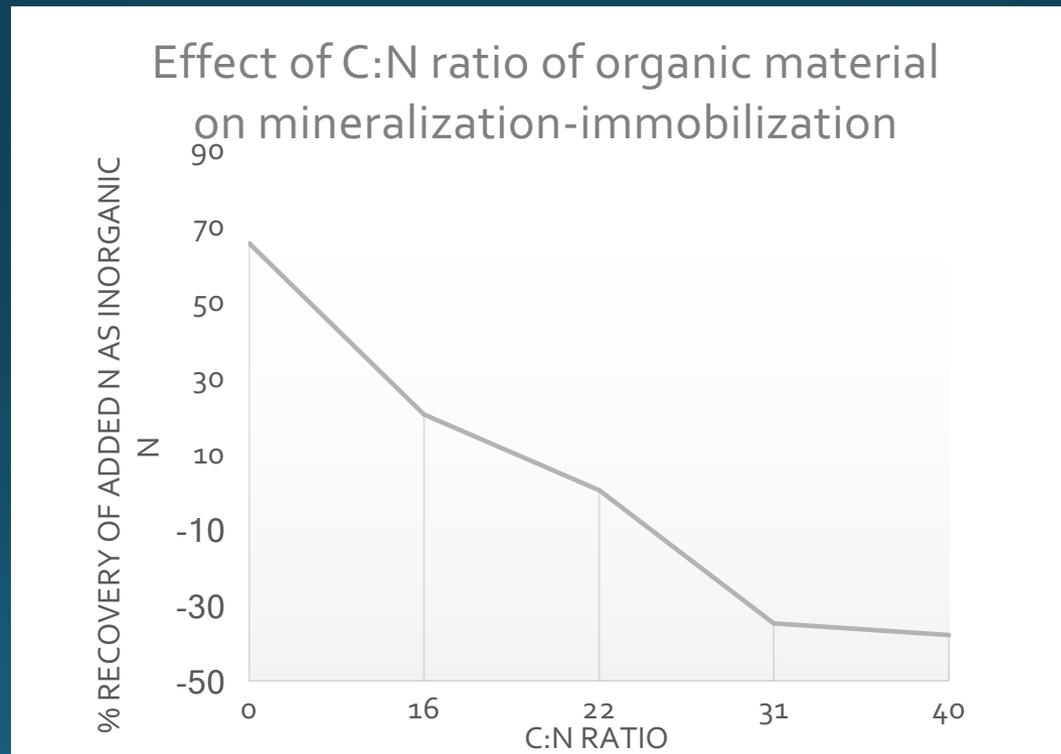
# N Mineralization

- Conversion of organic N to  $\text{NH}_4^+$  through two reactions:
  - **aminization:** converts proteins in residues to *amino acids, amines, and urea*
  - **ammonification:** further converts organic N compounds to inorganic  $\text{NH}_4^+$ 
    - The  $\text{NH}_4^+$  produced is subject to several fates:
      - Nitrification
      - N uptake
      - immobilization
      - $\text{NH}_4^+$  fixation
      - Volatilization

# N Immobilization

- Conversion of inorganic N ( $\text{NH}_4^+$  and  $\text{NO}_3^-$ ) to organic N
  - If decomposing residues contain low N, microorganisms will immobilize  $\text{NH}_4^+$  and  $\text{NO}_3^-$  in the soil solution

- C:N ratio



**TABLE 16-2** Validly Described Genera and Species of Root-Nodule Bacteria of Legumes. Changes to this taxonomy suggested by Young et al. (2001) are in bold type. Genera in the square brackets refer to better-known host legumes nodulated by each species of root-nodule bacteria. Common names are included for well-known legume genera. In several examples in this list, different species of root-nodule bacteria nodulate the same legume.\*

*Azorhizobium*\*\*

*A. caulinodans* [*Sesbania*]

*Bradyrhizobium*

*B. elkanii* [*Glycine*, soybean]

*B. japonicum* [*Glycine*]

*B. liaoningense* [*Glycine*]

*B. yuanmingense* [*Lespedeza*]

*Mesorhizobium*

*M. amorphae* [*Amorpha*]

*M. chacoense* [*Prosopis*, mesquite]

*M. ciceri* [*Cicer*, chickpea]

*M. huakuii* [*Astragalus*, milkvetch]

*M. loti* [*Lotus*]

*M. mediterraneum* [*Cicer*]

*M. plurifarium* [*Acacia*, *Leucaena*, *Ipil-ipil*]

*M. tianshanense* [*Glycyrrhiza*, *Sophora*]

*Rhizobium*

*R. etli* [*Phaseolus vulgaris*, bean]

*R. galegae* [*Galega*, *Leucaena*]

*R. gallicum* [*Phaseolus*, *Dalea*, *Onobrychis*, *Leucaena*]

*R. giardinii* [*Phaseolus*]

*R. hainanense* [*Stylosanthes*, *Centrosema*]

*R. huautlense* [*Sesbania*]

*R. indigoferae* [*Indigofera*]

*R. leguminosarum*

*bv trifolii* [*Trifolium*, clover]

*bv viciae* [*Pisum*, peas, *Vicia*, field beans, *Lathyrus*; and *Lens*, lentil]

*bv phaseoli* [*Phaseolus*]

*R. loessense* [*Astragalus*]

*R. mongolense* [*Medicago*, *Phaseolus*]

*R. sullae* [*Hedysarum*]

*R. tropici* [*Phaseolus*; *Leucaena*, *Dalea*, *Macroptilium*]

*Allorhizobium*

*A. undicola*, *R. undicola* [*Neptunia*]

*R. radiobacter* [non-nodulating saprophyte], *R. rhizogenes* [causes hairy root disease],

*R. rubi*, *R. vitis*

*Sinorhizobium*

*S. abri* [*Abrus*]

*S. americanus* [*Acacia*]

*S. arboris*

*S. fredii* [*Glycine*]

*S. indiaense* [*Sesbania*]

*S. kostiense*

*S. kummerowiae* [*Kummerowia*]

*S. medicae* [*Medicago*]

*S. meliloti* [*Melilotus*, sweetclover; *Medicago*, alfalfa; and *Trigonella*, fenugreek]

*S. morelense* [*Leucaena*]

*S. saheli*, *S. sahalense* [*Sesbania*]

*S. terangae* [*Sesbania*, *Acacia*]

*S. xinjiangense* [*Glycine*]

\*Other genus and species names exist in the literature. Some predate the present names; others have not been accepted as valid.

\*\*Strains which have not yet been recognized as belonging to any named species are usually identified by the host from which they were isolated—for example, *Rhizobium* spp. (*Acacia*) or *Bradyrhizobium* spp. (*Lupinus*).

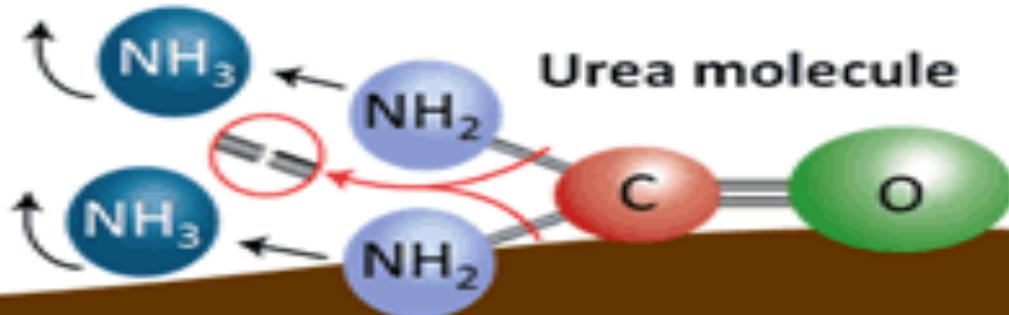
# Events Leading to Nodulation and Dinitrogen Fixation in Legumes

- Attachment of rhizobia to the root begins within 1 minute of inoculation.
- Number of attached rhizobia increases with time up to several hours.
- Root hair curling begins within 5 hours.
- Infection threads visible in the root hair within 3 days of inoculation.
- Nodule become visible within 5 to 12 days.
- N<sub>2</sub> fixation is often evident in 15 day-old plants.

# N use and fertility

- Global use of manufactured N fertilizers has increased by a factor of 8 over past six decades
- Efficiency of fertilizer N is frequently low
  - Often <50% of applied N is taken up by crop
  - For recommended rates 50-70% efficiency assumed

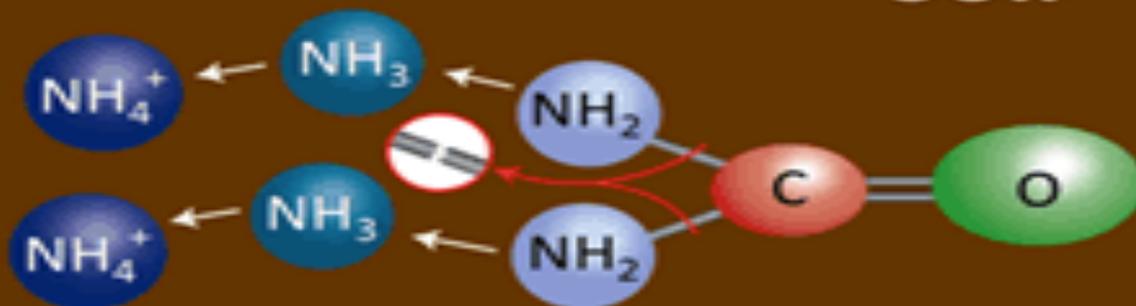
Gaseous ammonia released to the air



## Soil

If urea is hydrolyzed by urease at the soil surface, part of the NH<sub>3</sub> that is released may be lost to the air.

## Soil



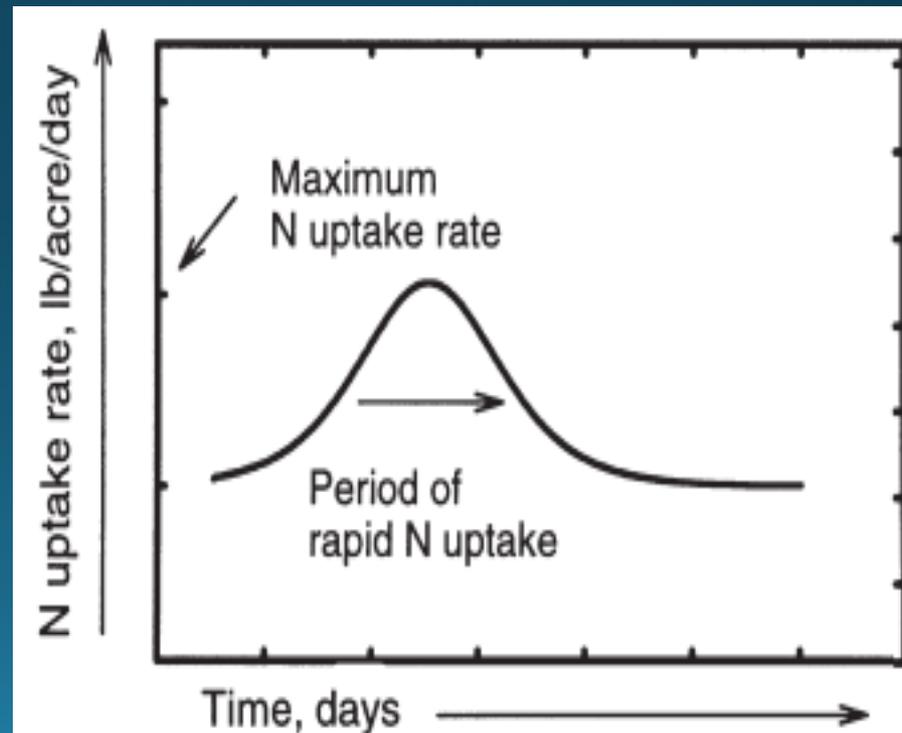
NH<sub>4</sub><sup>+</sup> binds to soil.

Urea molecule

If urea is hydrolyzed by urease within the soil, the NH<sub>3</sub> that is released reacts with soil water to form NH<sub>4</sub><sup>+</sup>, which adheres to soil components.

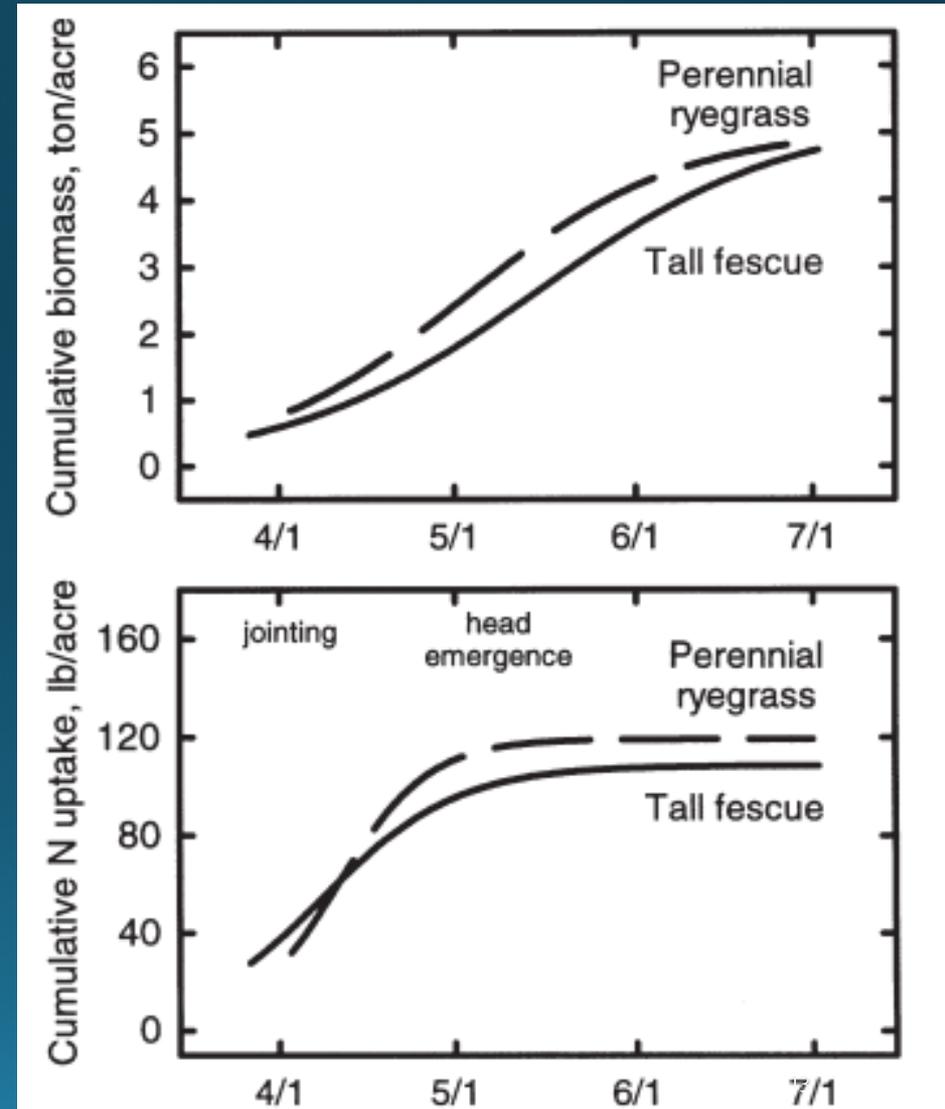
# Timing of N availability

- Seedling stage: N demand small
  - Rapid vegetative growth (4 to 6 weeks): High N demand
    - Little or no additional N uptake
  - Reproductive growth (flowering): N moves from leaves to seed
    - Little or no additional N uptake
- What time is most efficient for N fertilization?



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# cereal rye cover crop scavenges nitrate over the winter

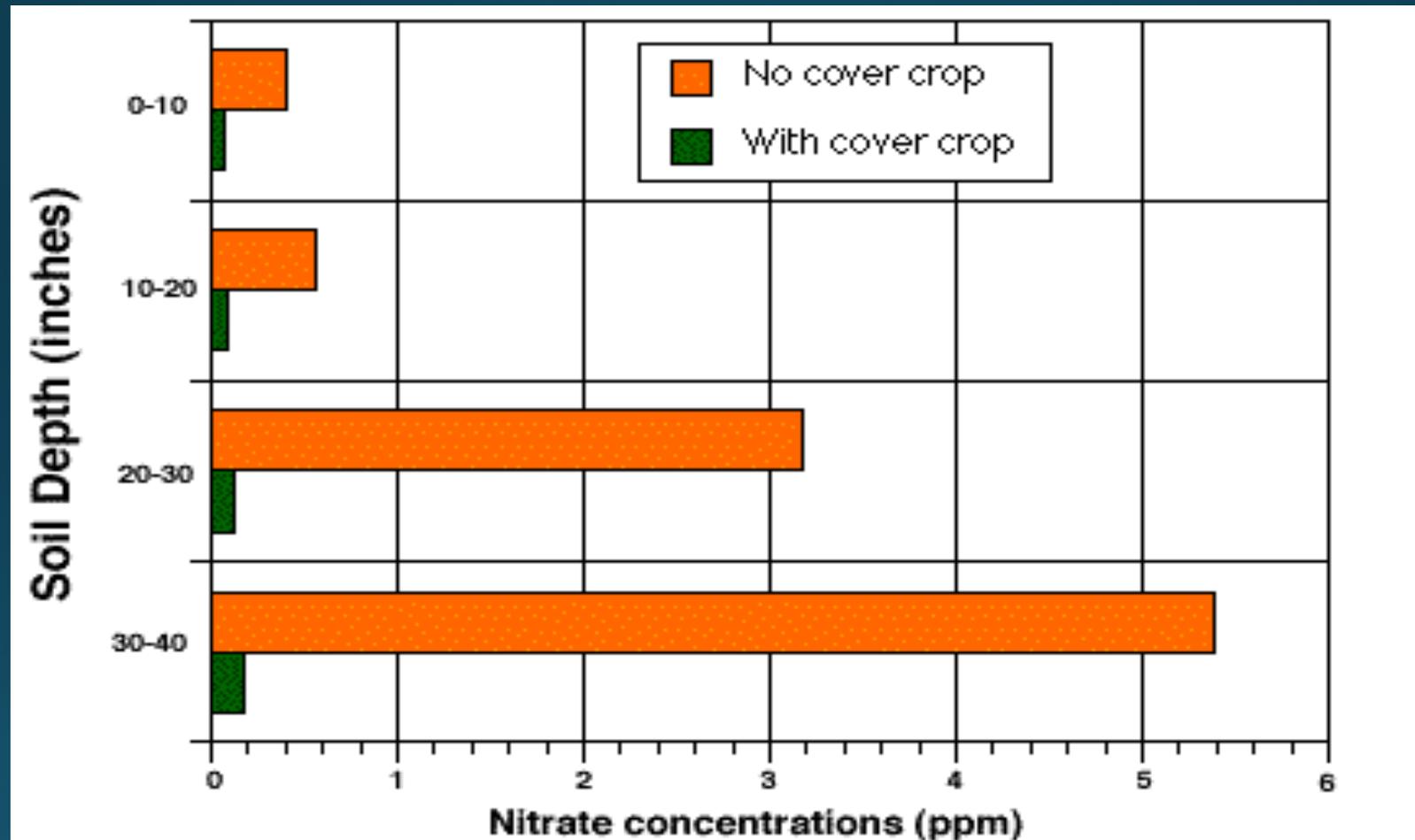


Figure 3. Effect of a cereal rye cover crop on soil nitrate concentrations (ppm) in broccoli plots fertilized the previous spring with 250 pounds N/acre. Samples were taken April 15, 1992.

(Data from Hemphill and Hart, 1993.)

# Part 2

# Estimated Average Rates of Biological N<sub>2</sub> Fixation

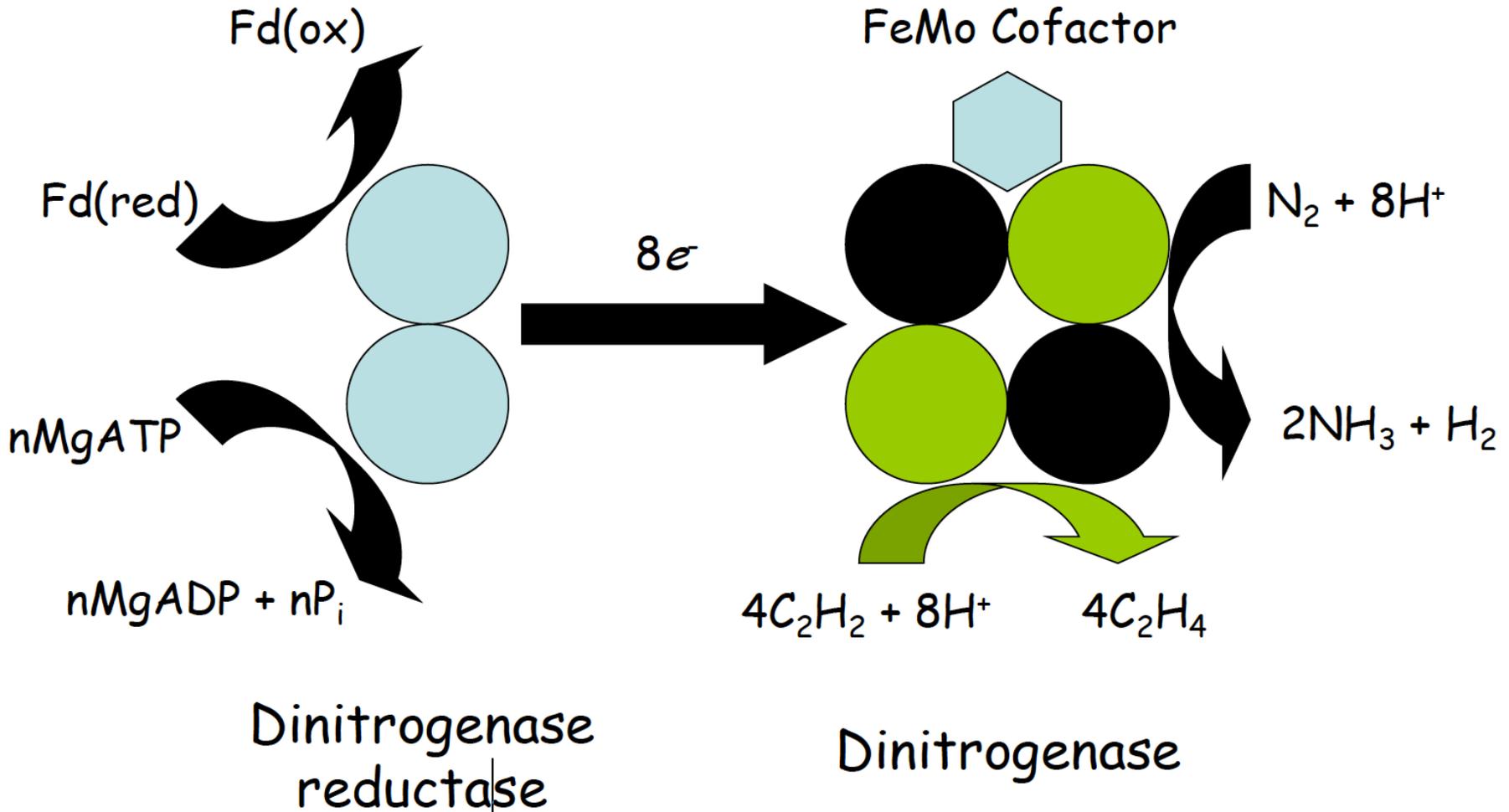
Organism or system	N <sub>2</sub> fixed (kg ha <sup>-1</sup> y <sup>-1</sup> )
<b>Free-living microorganisms</b>	
<i>Cyanobacteria</i>	25
<i>Azotobacter</i>	0.3
<i>Clostridium pasteurianum</i>	0.1-0.5
<b>Grass-Bacteria associative symbioses</b>	
<i>Azospirillum</i>	5-25
<b>Cyanobacterial associations</b>	
<i>Gunnera</i>	10-20
<i>Azolla</i>	50
Lichens	40-80
<b>Leguminous plant symbioses with rhizobia</b>	
Grain legumes ( <i>Glycine, Vigna, Lespedeza, Phaseolus</i> )	20-180
Pasture legumes ( <i>Trifolium, Medicago, Lupinus</i> )	50-300
<b>Actinorhizal plant symbioses with <i>Frankia</i></b>	
<i>Alnus</i>	40-300
<i>Hippophaë</i>	1-150
<i>Ceanothus</i>	1-50
<i>Coriaria</i>	50-150
<i>Casuarina</i>	50

# Nitrogen Fixation Process

## Energetics

- $\text{N}\equiv\text{N}$
- Haber-Bosch (100-200 atm, 400-500°C, ~1,900 kJ kg<sup>-1</sup> N)
- Nitrogenase (~950 kJ kg<sup>-1</sup> N)

# Nitrogenase



# Genetics of Nitrogenase

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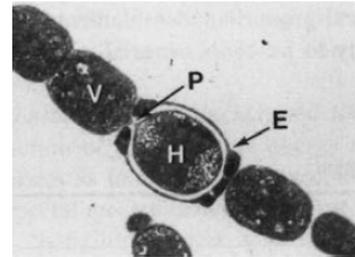
Gene	Properties and function
<i>nifH</i>	Dinitrogenase reductase
<i>nifDK</i>	Dinitrogenase
<i>nifA</i>	Regulatory, activator of most <i>nif</i> and <i>fix</i> genes
<i>nifB</i>	FeMo cofactor biosynthesis
<i>nifEN</i>	FeMo cofactor biosynthesis
<i>nifS</i>	Unknown
<i>fixABCX</i>	Electron transfer
<i>fixK</i>	Regulatory
<i>fixLJ</i>	Regulatory, two-component sensor/effector
<i>fixNOQP</i>	Electron transfer
<i>fixGHIS</i>	Transmembrane complex

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# Types of Biological Nitrogen Fixation

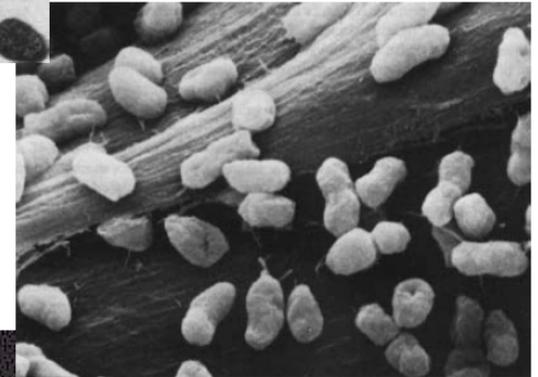
## Free-living (asymbiotic)

- Cyanobacteria
- *Azotobacter*



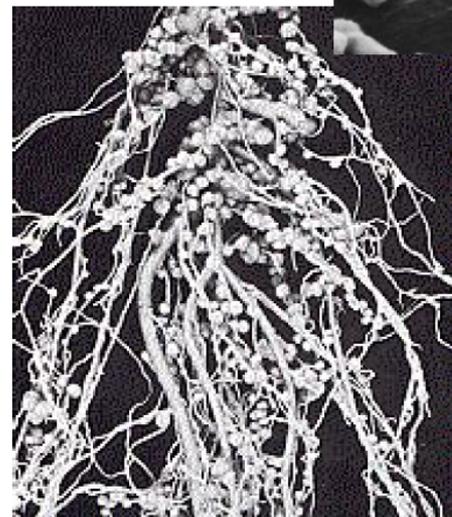
## Associative

- Rhizosphere-*Azospirillum*
- Lichens-cyanobacteria
- Leaf nodules



## Symbiotic

- Legume-rhizobia
- Actinorhizal-*Frankia*



# Free-living N<sub>2</sub> Fixation

## Energy

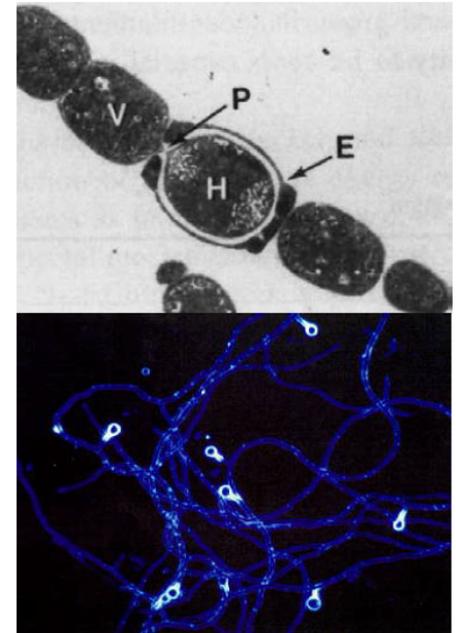
- 20-120 g C used to fix 1 g N

## Combined Nitrogen

- *nif* genes tightly regulated
- Inhibited at low NH<sub>4</sub><sup>+</sup> and NO<sub>3</sub><sup>-</sup> (1 μg g<sup>-1</sup> soil, 300 μM)

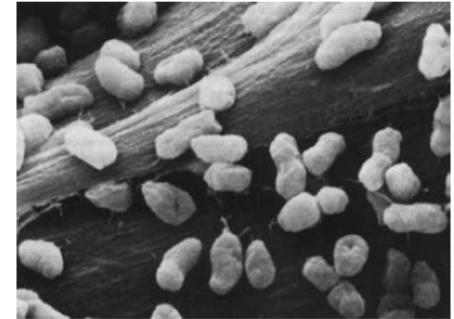
## Oxygen

- Avoidance (anaerobes)
- Microaerophilly
- Respiratory protection
- Specialized cells (heterocysts, vesicles)
- Spatial/temporal separation
- Conformational protection



# Associative N<sub>2</sub> Fixation

- Phyllosphere or rhizosphere (tropical grasses)
- *Azospirillum*, *Gluconacetobacter*
- 1 to 10% of rhizosphere population
- Some establish within root
- Same energy and oxygen limitations as free-living
- *Gluconacetobacter diazotrophicus* lives in internal tissue of sugar cane, grows in 30% sucrose, can reach populations of 10<sup>6</sup> to 10<sup>7</sup> cells g<sup>-1</sup> tissue, and fix 100 to 150 kg N ha<sup>-1</sup> y<sup>-1</sup>



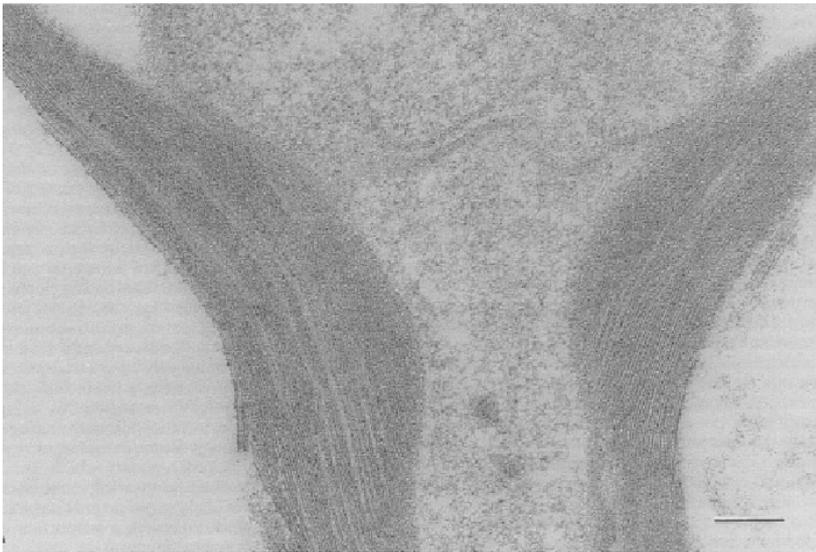
# Phototrophic N<sub>2</sub>-fixing Associations

- Lichens-cyanobacteria and fungi
- Mosses and liverworts — some have associated cyanobacteria
- *Azolla-Anabaena (Nostoc)* — cyanobacteria in stem of water fern
- *Gunnera-Nostoc* — cyanobacteria in stem nodule of dicot
- *Cycas-Nostoc* — cyanobacteria in roots of gymnosperm



# *Frankia* and Actinorhizal Plants

- Actinobacteria (Gram +, filamentous); septate hyphae; spores in sporangia; thick-walled vesicles



*Frankia* vesicles showing thick walls that confer protection from oxygen. Bars are 100 nm.



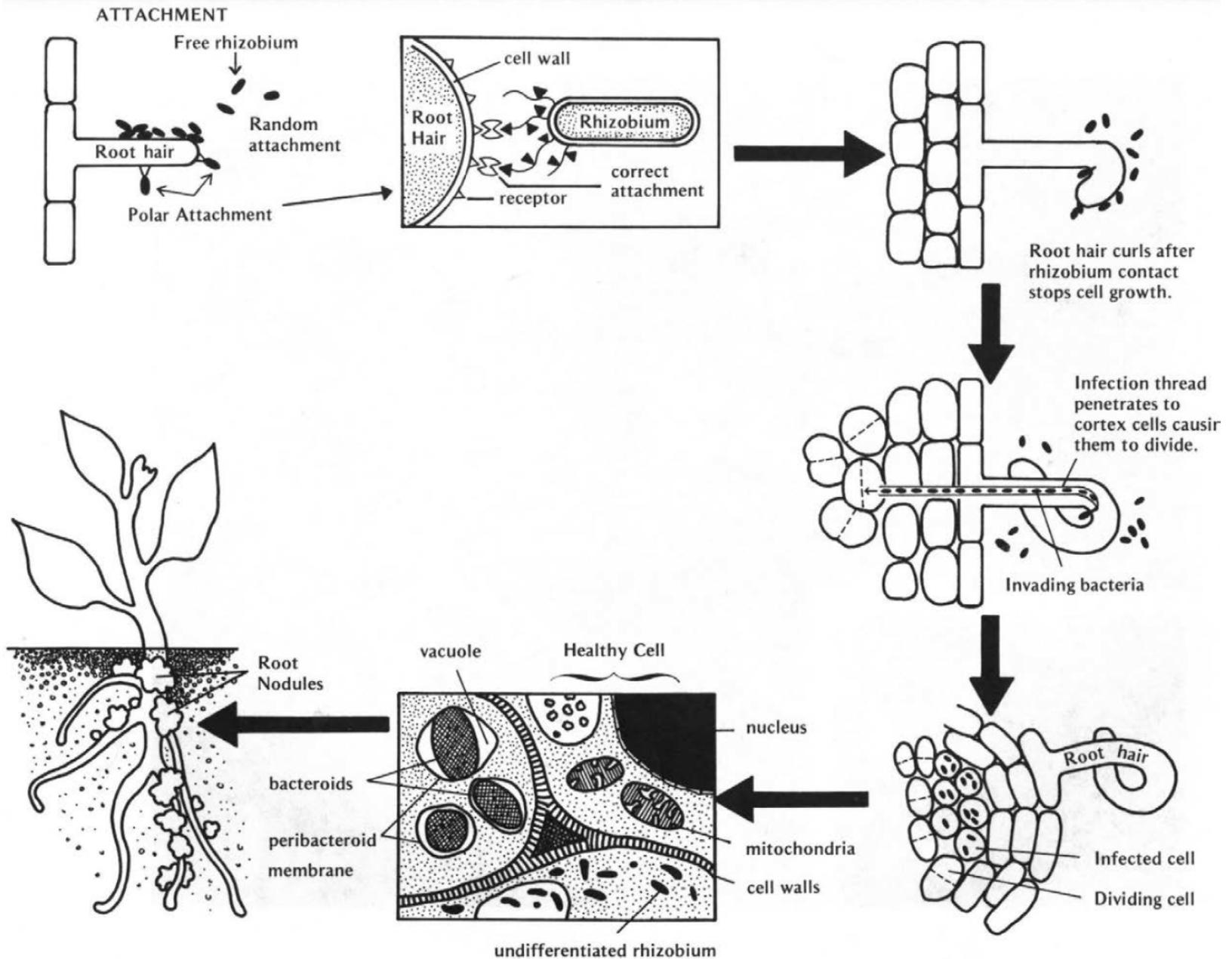
# Actinorhizal Plant Hosts

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Family	Genera
Betulaceae	<i>Alnus</i>
Casuarinaceae	<i>Allocasuarina, Casuarina, Ceuthostoma, Gymnostoma</i>
Myricaceae	<i>Comptonia, Myrica</i>
Elaeagnaceae	<i>Elaeagnus, Hippophaë, Shepherdia</i>
Rhamnaceae	<i>Ceanothus, Colletia, Discaria, Kentrothamnus, Retanilla, Talguenea, Trevoa</i>
Rosaceae	<i>Cercocarpus, Chamaebatia, Cowania, Dryas, Purshia</i>
Coriariaceae	<i>Coriaria</i>
Datisceae	<i>Datisca</i>

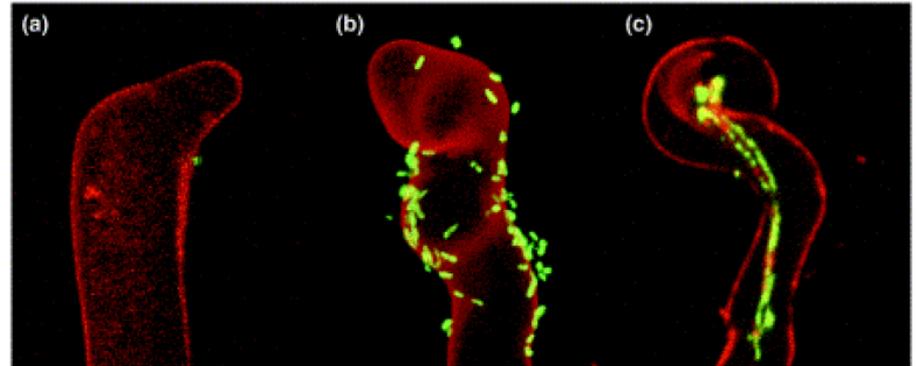
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# Nodulation in Legumes

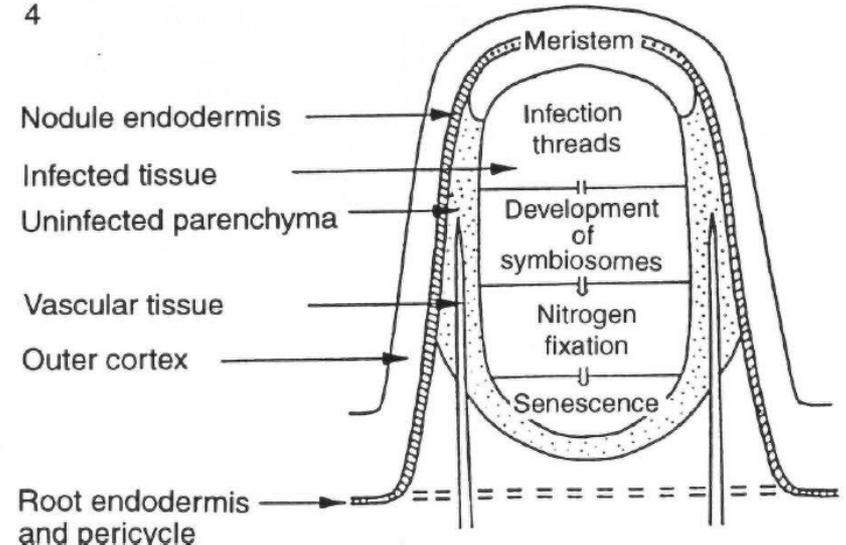


# Infection Process

- Attachment
- Root hair curling
- Localized cell wall degradation
- Infection thread
- Cortical cell differentiation
- Rhizobia released into cytoplasm (symbiosome formation)
- Bacteroid differentiation
- Induction of nodulins



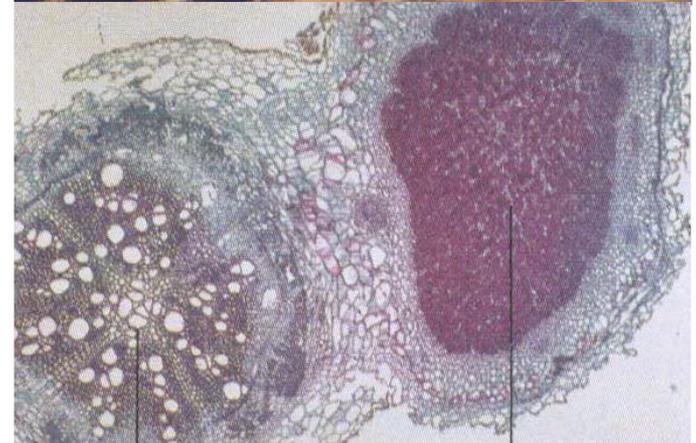
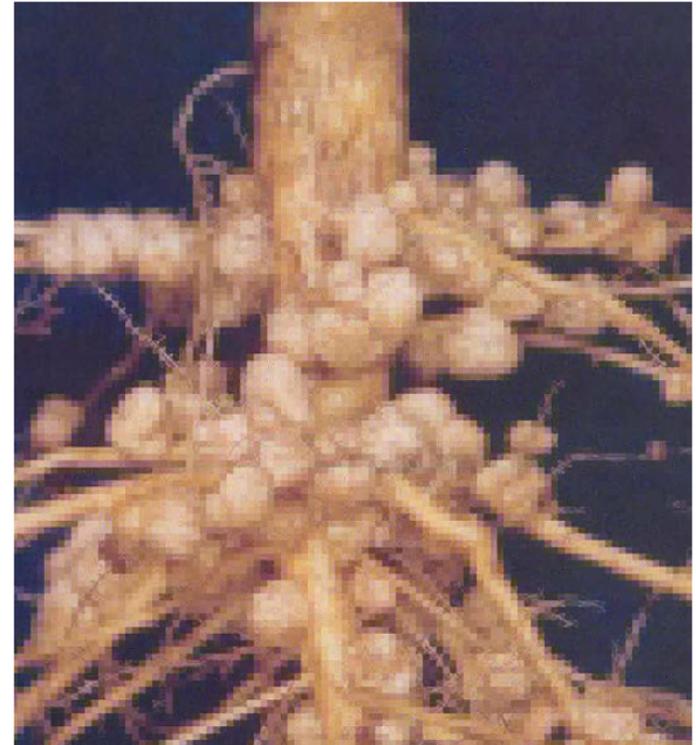
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Indeterminate (pea) nodule

# Legume-Rhizobium Symbiosis

- The subfamilies of legumes (Caesalpinioideae, Mimosoideae, Papilionoideae), 700 genera, and 19,700 species of legumes
- Only about 15% of the species have been evaluated for nodulation
- *Rhizobium*
  - Gram -, rod
  - Most studied symbiotic N<sub>2</sub>-fixing bacteria
  - Now subdivided into several genera
  - Many genes known that are involved in nodulation (*nod*, *nod*, *noe* genes)



# Taxonomy of Rhizobia

Genus	Species	Host plant
<i>Rhizobium</i> (~30 species)	<i>leguminosarum</i> bv. <i>trifolii</i>	<i>Trifolium</i> (clovers)
	"	<i>Pisum</i> (peas), <i>Vicia</i> (field beans), <i>Lens</i> (lentils)
	"	<i>Phaseolus</i> (bean), <i>Leucaena</i>
	<i>tropici</i>	<i>Phaseolus</i> (bean)
	<i>etli</i>	<i>Phaseolus</i> (bean)
	<i>galegae</i>	<i>Galegae</i> (goat's rue)
<i>Ensifer</i> ( <i>Sinorhizobium</i> , ~10 species)	<i>meliloti</i>	<i>Medicago</i> (alfalfa)
	<i>fredii</i>	<i>Glycine</i> (soybean)
	<i>saheli</i>	<i>Sesbania</i>
	<i>americanum</i>	<i>Acacia</i>
<i>Bradyrhizobium</i> (~13 species)	<i>japonicum</i>	<i>Glycine</i> (soybean)
	<i>elkanii</i>	<i>Glycine</i> (soybean)
	<i>liaoningense</i>	<i>Glycine</i> (soybean)
<i>Azorhizobium</i> (2 species)	<i>caulinodans</i>	<i>Sesbania</i> (stem nodule)
<i>Mesorhizobium</i> (~20 species)	<i>loti</i>	<i>Lotus</i> (trefoil)
	<i>huakuui</i>	<i>Astragalus</i> (milkvetch)
	<i>ciceri</i>	<i>Cicer</i> (chickpea)
	<i>mediterraneum</i>	<i>Cicer</i> (chickpea)
<i>Allorhizobium</i>	<i>undicola</i>	<i>Neptunia</i>
<i>Photrhizobium</i>	spp.	<i>Aeschynomene</i> (stem nodule)